

# The Effect of *Bifidobacterium Longum* on Expression of *Hfq* in *Escherichia Coli* O157:H7

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## ABSTRACT

**Background and Aim:** *Escherichia coli* (*E. coli*) O157:H7 is the primary cause of food poisoning and urinary tract infection via Shiga toxin. The main transcription factor for Shiga toxin is *Hfq* gene. It is reported that probiotics can reduce the production of toxin substances in pathogenic bacteria. This study was aimed to investigate the influence of *Bifidobacterium Longum* (*B. Longum*) on the expression level of the *Hfq* gene as the main virulence factor in *E. coli* O157:H7.

**Materials and Methods:** *Bifidobacterium Longum* was cultured in MRS (De Man, Rogosa, and Sharpe) medium. Different concentrations of *B. Longum* was co-cultured in Mueller-Hinton Agar (MHA) along with *E. coli* in different time points. Total RNA was extracted followed by synthesizing cDNA. The *Hfq* gene expression was evaluated by SYBR Green real-time PCR. The 16S ribosomal RNA was used as an internal control for data normalization.

**Results:** The results showed reduction in *E. coli* growth at different co-culture time points with *B. Longum*. Molecular analysis demonstrated that *B. Longum* has the ability to significantly reduce the expression of *Hfq* virulence gene ( $P=0.0023$ ).

**Conclusion:** Probiotic bacteria inhibit the growth of pathogenic bacteria by producing bacteriocins and reducing the expression level of the virulence genes. They can also play role in inhibiting intoxications and infections caused by *E. coli*.

**Keywords:** *Bifidobacterium Longum*, *E. coli*, *Hfq* gene, Probiotics

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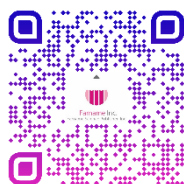
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## 1. Introduction

*Escherichia coli* (*E. coli*) is a Gram-negative bacterium with certain serotypes, such as O157:H7, that can cause foodborne illness and diarrhea in human (1). This bacterium is the most common cause of urinary tract infections, accounting for approximately 90% of the urinary infections in women (2). At the same time, this important pathogenic

microorganism has a variety of uses in industry and medicine (1). Based on the toxins production there are different types of *E. coli* including Shiga toxin-producing *E. coli* (STEC), verotoxin-producing *E. coli* (VTEC), and enterohemorrhagic *E. coli* (EHEC). including O157-, CFT073-, and K-12-specific (2). One of the highly important serotypes of EHEC is *E. coli*

O157:H7, which is transmitted to human through food and causes dangerous complications such as bloody colitis (3). *E. coli* expresses the *Hfq* gene as regulator of the virulence traits (4).

The *Hfq* in *E. coli* is a very conserved protein modulating the degradation of target RNAs, facilitating the sRNA-mRNA pairing, and stabilizing the sRNAs (3). Recently, scientists focused on recruiting the microbiome as a novel therapeutic for the *E. coli*-related infections (3, 5).

*Bifidobacterium* is the most important anaerobic Gram-positive, highly pleomorphic, and non-spore-forming probiotic bacteria in the normal flora of intestine (6). This bacterium has beneficial effects including the production of essential vitamins and increased digestibility of proteins as well as preventing intestinal infections and diarrhea, reducing radiotherapy-induced disorders, treating certain brain lesions, lowering blood cholesterol levels, reducing cancer-related compounds, and immunological effects by strengthening the immune system or reducing the lactose intolerance effects (7, 8).

In 2008, Carey et al (9) reported the impact of probiotics on reducing the production of Shiga toxin type 2 (STX2) in *E. coli* O157:H7. Ahmadizadeh et al (10), in 2018, documented the effect of probiotics in reducing the expression of verotoxin genes *STX1* and *STX2*. Takahashi et al (11), in 2004 reduced the production of Shiga toxins *STX1* and *STX2* in *E. coli*-infected mice using the probiotic bacteria *Clostridium butyricum*. Asahara et al (12) reported in 2004 that probiotics could reduce the mortality from STEC O157:H7 in mice. Wan et al (13), in 2019 suggested using probiotics to control foodborne bacteria and toxin-producing contaminants, proposing that their addition can control these factors. Cordonnier et al (14) suggested that probiotics and enterohemorrhagic *Escherichia coli* has an effective strategy against a deadly enemy. Ku et al (15) recently published an article stating that probiotic consumption is very safe for humans and can be used to combat the microbial infections, especially toxin-producing microbes.

The aim of this research was to study the effect of co-cultivation of the *Bifidobacterium Longum* (*B. Longum*) as a probiotic bacterium with pathogenic *E. coli* O157:H7 on the expression level of the virulence *Hfq* gene and its growth rate.

## 2. Materials and Methods

### 2.1. Materials

*E. coli* O157:H7 bacteria were obtained from the Collection Center of Infectious and Industrial

Microorganisms, (Iran) and cultured in liquid LB medium for 16 hours at 37°C. The cultured sample was then stocked in 25% glycerol solution and stored at -70°C.

The culture medium, necessary chemicals, and solutions used in this research include: DEPC, diethyl pyrocarbonate; dNTPs, deoxynucleotide triphosphate monomers; EDTA, ethylenediaminetetraacetic acid; HCL, hydrochloric acid; KCl, potassium chloride; KH<sub>2</sub>PO<sub>4</sub>, potassium dihydrogen phosphate; Na<sub>2</sub>HPO<sub>4</sub>, disodium hydrogen phosphate; NaCl, sodium chloride; NaHCO<sub>3</sub>, sodium bicarbonate; NaOH, sodium hydroxide; and phosphate buffer saline (PBS).

### 2.2. Methods

#### 2.2.1. *E. coli Hfq* Gene PCR Amplification

The salting-out method was employed for the DNA extraction of *E. coli*. Briefly, the cultured bacteria were centrifuged at 10,000 rpm for 5 min and the supernatant was mixed with 850 µl pre-heated lysing buffer at 65°C. The samples were then incubated at 65°C for 30 min and 5-6 min, respectively. Then, 700 µl chloroform-isoamyl alcohol solution was added to the samples and centrifuged at 10,000 rpm for 10 min. The isolated DNA was then washed with cold isopropanol (×1.5 volume of the sample), stored overnight at -70°C, centrifuged at 10,000 rpm for 5 min, and dried at room temperature. Finally, DNA samples were dissolved in 50 µl DNase-free deionized distilled water and stored until further evaluations.

Then, polymerase chain reaction (PCR) was performed mixing 12.5 µl PCR master mix (2x), 100 ng DNA, specific primers, and H<sub>2</sub>O up to 25 µl volume. The PCR reaction cycling involved initial denaturation at 95°C for 4 min, followed by 32 cycles of denaturation at 94°C for 1 min, primer annealing at 57°C for 80 sec, extension at 72°C for 1 min, and a final extension step at 72°C for 10 min. The amplified DNA samples were also loaded on 1% agarose gel with loading buffer in a 6:1 ratio and evaluated using SYBR Green dye and UV light.

#### 2.2.2. Co-culturing *E. coli* with *Bifidobacterium*

Initially, *E. coli* was cultured in Mueller-Hinton Agar (MHA) under aerobic conditions. Simultaneously, *Bifidobacterium* was cultured in MRS (De Man, Rogosa, and Sharpe) under anaerobic conditions completely deoxygenating by purging with nitrogen gas and autoclaved. Then, different concentrations of this bacterium were used to treat *E. coli* at concentration of 0.5 McFarland. To investigate the expression of the virulence gene, the cultures were collected and centrifuged at various time intervals. The range of concentrations were obtained from

preliminary experiments (serial dilutions) and literature (7, 8).

### 2.2.3. The Hfq Expression Levels Evaluation

For RNA extraction, 500 µl Trizol solution was added to the culture medium and incubated at 45°C for 90 min. After that, 200 µl chloroform was added, incubated at room temperature for 5 min, and centrifuged at 12,000 rpm for 10 min. The supernatant was mixed with 2.5x cold isopropanol and stored at -70°C overnight. Then, samples were centrifuged at 12,000 rpm for 10 min and dried. Then, 20 µl DEPC-treated water was added to the RNA pellet and incubated for 10 min at room temperature. The OD was measured in ng/ml using a NanoDrop instrument and the samples were stored at -70°C.

For the reverse transcription and cDNA synthesis, a mixture of 500 ng total RNA, 0.2 µM random hexamer primer, 1 µl dNTP, and DEPC-treated water were

incubated at 65°C for 5 min. Then, 5 units of MMLV enzyme, 1x buffer for MMLV RT, and 1 U/µl RNase inhibitor were added and incubated at 25°C for 10 min, 42°C for 60 min, and 72°C for 10 min.

Negative control (RNA sample without reverse transcriptase) and positive control (known concentration of target RNA) were used during RNA extraction and cDNA synthesis.

In real-time PCR reactions, cDNA, master mix, forward and reverse primers (0.2 µM/each) and DEPC-treated water were mixed and the cycling was performed as mentioned before. The primers were designed using the oligo7 program, blasted through the NCBI website ([www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)) and synthesized by Takapouzist Company, Iran. The 16S ribosomal RNA was used as an internal control for data normalization. The specifications of the used primers are shown in Table 1.

**Table 1.** Specifications of primers used in real-time PCR method

Gene	The length of the pieces	Primer sequence	Annealing temperature
Hfq	180 bp	Forward: 5'-ATAAATCGCCATTCGTTGACTA-3'	51 °C
		Reverse: 5'-AGAAGGCCCACTGAGATCATC-3'	
16s rDNA	90 bp	Forward: 5'-ACTCTGTTATTAGGGAAGAA-3'	53 °C
		Reverse: 5'-AACGCTTGCCACCTACGTAT-3'	

### 2.2.4. Data Analysis

The cycle threshold (CT) values were used to evaluate the expression level of Hfq gene. The obtained data were normalized to 16s rDNA housekeeping gene and the gene expression ratio was calculated using equation provided by Asahara (12):

$$\text{Gene expression ratio} = \frac{(E_{\text{target}})^{\Delta CP_{\text{target}}(\text{control-sample})}}{(E_{\text{ref}})^{\Delta CP_{\text{ref}}(\text{control-sample})}} \quad (1)$$

The relative expression ratio of a target gene is computed, based on its real-time PCR efficiencies (E) and the crossing point (CP) difference (Δ) of an unknown sample versus a control (ΔCP<sub>control</sub> – sample).

The data analysis was performed via SPSS software. Data normality was checked via the Shapiro-Wilk test and due to the normal distribution, the comparison between two groups was performed via independent

t-test. The P-value less than 0.05 was considered significant.

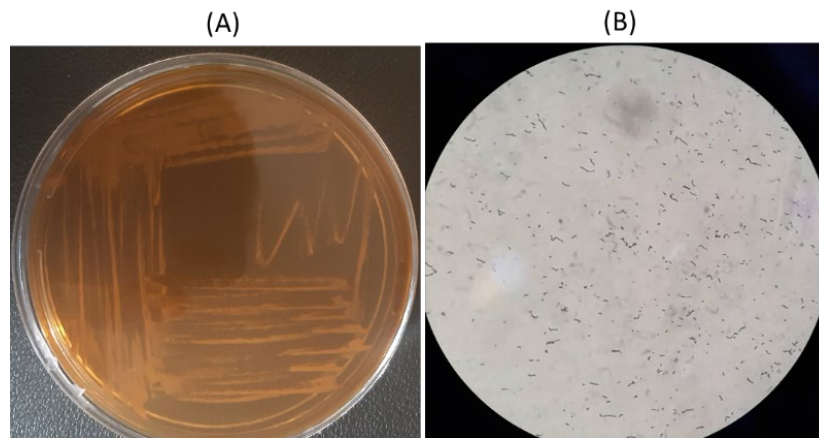
## 3. Results

### 3.1. Bifidobacterium Longum Culture Results

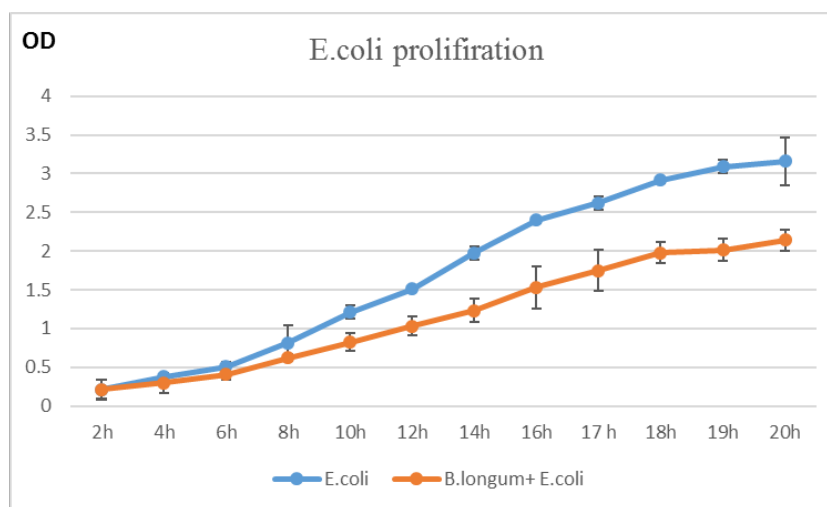
The culture of B. Longum was conducted in anaerobic conditions and in MRS shown in Figure 1.

### 3.2. Co-Culture of E. coli with B. Longum

The growth of E. coli alone and in the presence of B. Longum was conducted at 37°C and aerobic conditions over 20 hours using 1 ml of bacteria in the cultured medium. Following each corresponding incubation interval, the culture was collected, centrifuged at 300 rpm and the bacterial pellet was resuspended in 1 ml PBS and growth rate was measured using Nanodrop at 600 nm wavelength in triplicates (Figure 2). B. Longum showed the capability to inhibit the growth of E. coli significantly, achieving nearly a two-fold reduction.



**Figure 1.** Cultivation of *B. Longum* in MRS (De Man, Rogosa, and Sharpe). The culture was observed with naked eye (A) and (B) under the microscope (10x).

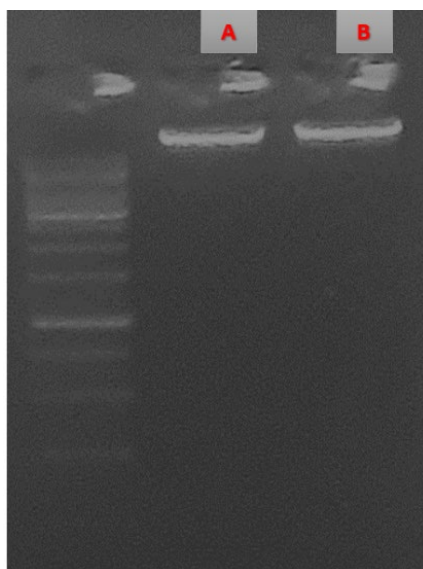


**Figure 2.** *E. coli* proliferation rates alone and in co-culture with *B. Longum* based on OD at 600 nm at 2, 4, 6, 8, 10, 12, 14, 16, 17, 18, 19, and 20 hours.

### 3.3. Confirmation of *Hfq* Gene Presence in *E. coli*

The presence of the *Hfq* gene in *E. coli* was confirmed for further comparative gene expression analysis. The DNA extraction from *E. coli* bacteria and

the confirmation of the presence of the *Hfq* gene were carried out before and after DNA amplification of *E. coli* O157:H7 (Figure 3). After performing PCR, a distinct amplification band for the *Hfq* gene of approximately 180 bp was observed as expected.



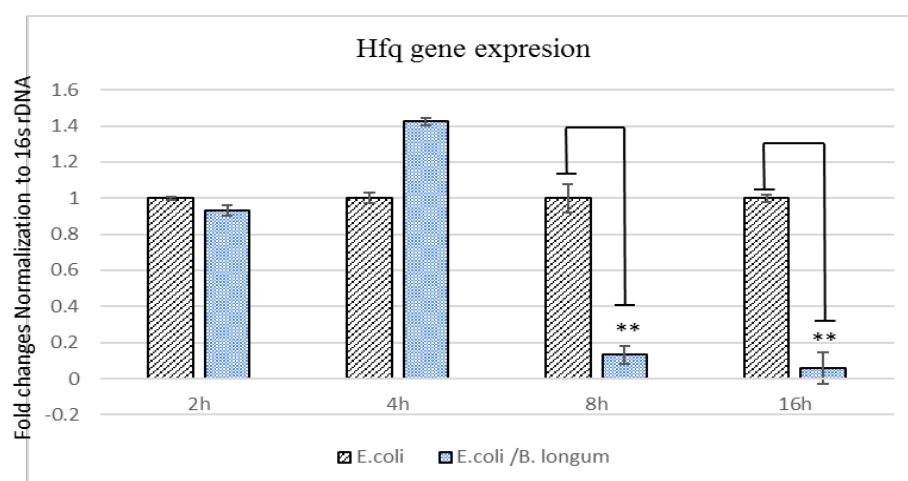
**Figure 3.** Gel electrophoresis of *E. coli* extracted DNA (*Hfq* gene) on 1% agarose gel before (A) and after (B) DNA amplification.

### 3.4. The *Hfq* Expression Levels

To evaluate the changes in the expression of the *Hfq* gene, total RNA extracted from *E. coli* bacteria alone or in co-culture with *B. Longum* was evaluated.

The obtained results from evaluating the expression levels of *Hfq* showed that *B. Longum* could inhibit *E.*

*coli* growth at 8 and 16 hours after co-culturing compared to the cultured *E. coli* alone ( $P= 0.0023$ ). However, the observed changes at 2 and 4 hours after co-culturing was not statistically significant ( $P>0.05$ ). The results were compared using fold-change assessment of expression levels (Figure 4).



**Figure 4.** Expression levels of *Hfq* gene in *E. coli* alone and in co-culture with *B. Longum* at 2, 4, 8, and 16 hours. The results of gene expression were normalized to 16S rDNA. \*\* shows significant level at  $P<0.001$ .

## 4. Discussion

The results of the current study demonstrated that probiotic *B. Longum* not only inhibited the growth of *E. coli* but also significantly reduced the expression of *Hfq* virulence gene. Notably, the reduction in *E. coli* growth is attributed not only to the production of bacteriocin by *B. Longum* but also to its impact on reducing the expression of the *Hfq* gene. It has been shown that reduction in the expression of this

virulence gene may be attributed to the subsequent reduction in the growth of pathogenic bacteria.

Investigating the resistance and virulence of pathogenic bacteria is essential and the World Health Organization (WHO) has recently emphasized for monitoring *E. coli* O157:H7 (16-19). Therefore, the development of innovative therapeutic approaches to control this infection is crucial, and the use of



probiotics has gained significant attention in the past two decades (20-24).

The use of antibiotics for the treatment and prevention of infections caused by *E. coli* has not only led to the development of drug resistance but also has disrupted the natural balance of the beneficial normal flora in the digestive system, making the body susceptible to the various intestinal diseases (25-28). In contrast, today, the use of probiotic bacteria, which are symbiotic and beneficial microorganisms that combat pathogenic microbes in the living environment, can immunize individuals against disease-causing agents through dietary means (29-31).

Probiotics are live microorganisms consumed as dietary supplements or with fermented dairy products. Their consumption plays a significant role in various parts of the body, including mouth, digestive system, urinary and reproductive system, and upper respiratory system for preventing infections (32-34).

Probiotics include various strains of *Lactobacillus*, *Saccharomyces*, *Bifidobacterium*, *Enterococcus*, and *Clostridium* that can be used for both the treatment and prevention of infectious diseases. These probiotics perform through various mechanisms such as antimicrobial substances, organic acids, activation of the body immune system, complement system, and competition for the nutrients with microbial pathogens. They inhibit the growth and proliferation of pathogenic agents in that specific area of the body (35-37).

Numerous studies have investigated the impact of probiotics such as *Bifidobacterium angulatum*, *Bifidobacterium bifidum*, *Lactobacillus acidophilus*, and *Lactobacillus casei* on the growth rate of *E. coli* under simultaneous growth conditions. A systematic review by Torabian et al (35) demonstrated that probiotics could inhibit bacterial virulence factors.

Our study uniquely identifies the reduction in *Hfq* gene expression as a key mechanism through which *Bifidobacterium* impacts *E. coli* O157:H7, a mechanism not extensively explored in previous research. Antibiotics have also been employed for this purpose, and several notable studies have been conducted in this regard (9, 16).

What makes the current research unique is examination of the mechanism behind reducing the virulence of the pathogenic bacterium *E. coli*, particularly the O157:H7 strain. The *Hfq* gene is influential in the replication of the phage in *E. coli*. It is noteworthy to mention that although this gene plays a role in the replication of the pathogenic bacterium, it contributes to its virulence as well. The results of this research confirm these reports, as our findings indicate that *Bifidobacterium* not only inhibited the expression of the *Hfq* gene but also significantly reduced the

growth rate of *E. coli* in adjacent cultures at different time points. The *Hfq* protein is a key RNA-binding protein that plays a crucial role in regulating the stress response and virulence gene expression in bacteria. The mechanism of action of the *Hfq* protein is believed to prevent the silencing of the virulence genes by binding to the certain microorganisms, leading to increased expression of virulence genes such as *STX1* and *STX2* (4). The use of probiotic bacteria can play a protective role in this context (38-41).

## 5. Conclusion

This study demonstrated that *B. Longum* has a significant inhibitory effect on the growth and virulence of *E. coli*, particularly by affecting the expression of the *Hfq* gene. The experiments showed that co-culture of *E. coli* with *B. Longum* led to a marked decrease in *E. coli* proliferation, as evidenced by a two-fold reduction in bacterial growth under aerobic conditions at 37°C. This growth suppression was associated with the presence of bacteriocins produced by *B. Longum*, which are known to have antimicrobial properties. Further molecular analysis revealed a significant reduction in the expression levels of the *Hfq* gene in *E. coli* when co-cultured with *B. Longum* compared to the control. The downregulation of *Hfq* in *E. coli* suggests that *B. Longum* may interfere with the regulatory pathways controlled by this protein, thereby diminishing the pathogenic potential of *E. coli*.

The findings highlight the potential of *B. Longum* as a probiotic agent that can not only inhibit the growth of pathogenic bacteria through direct antimicrobial substances but also modulate bacterial gene expression, specifically targeting virulence factors like *Hfq*. This dual mechanism of physical inhibition of growth and molecular modulation of virulence underscores the therapeutic potential of *B. Longum* in preventing and managing the infections caused by pathogenic *E. coli* strains. This could pave the way for the development of novel probiotic therapies aimed at reducing bacterial infections and their associated complications. In the future studies, it is suggested to investigate the exact molecular interactions between *B. Longum* and *Hfq* at the genomic and proteomic levels. Moreover, exploring the potential of *B. Longum* as a probiotic therapy is important to mitigate infections caused by *E. coli* O157:H7. It is better to assess the impact of *B. Longum* on other virulence factors regulated by *Hfq* to provide a comprehensive understanding of its therapeutic potential.

## 6. Declarations

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### Ethical Considerations

None.

### Authors' Contributions

Sobhan Aboulhassanzadeh, Samin Aboulhassanzadeh, Zahra Nikkhooy: data collection and analysis. Sobhan Aboulhassanzadeh, Hamed Aghazadeh, and Behboud Jafari: writing manuscript.

### Conflict of Interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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### Consent to participate

Not applicable.

### Consent for publication

All authors reviewed and approved the final version of the manuscript.

### Availability of data and materials

All relevant data can be found within the manuscript.

### Supplementary information

The online version contains supplementary material.

## References

- Shringi S, Sheng H, Potter AA, Minnich SA, Hovde CJ, Besser TE. Repeated Oral Vaccination of Cattle with Shiga Toxin-Negative *Escherichia coli* O157:H7 Reduces Carriage of Wild-Type *E. coli* O157:H7 after Challenge. *Appl Environ Microbiol*. 2021;87(2):e02183-20. [DOI:10.1128/AEM.02183-20] [PMID] [PMCID]
- Moxley RA, Bargar TW, Kachman SD, Baker DR, Francis DH. Intimate Attachment of *Escherichia coli* O157:H7 to Urinary Bladder Epithelium in the Gnotobiotic Piglet Model. *Microorganisms*. 2020; 8(2):263. [PMID] [PMCID] [DOI:10.3390/microorganisms8020263]
- Dai J, Li C, Cui H, Lin L. Unraveling the anti-bacterial mechanism of *Litsea cubeba* essential oil against *E. coli* O157:H7 and its application in vegetable juices. *Int J Food Microbiol*. 2021;338: 108989. [DOI:10.1016/j.ijfoodmicro.2020.108989] [PMID]
- Kendall MM, Gruber CC, Rasko DA, Hughes DT, Sperandio V. *Hfq* virulence regulation in enterohemorrhagic *Escherichia coli* O157:H7 strain 86-24. *J Bacteriol*. 2011;193(24):6843-51. [DOI:10.1128/JB.06141-11] [PMID] [PMCID]
- Paritala V. Comparative gut Microbiome Evaluation of two Distinct Organisms: A Critical Review. *J Agric Sci Eng*. 2022;4(2):117-26.
- Guimarães J, Fasura Balthazar C, Silva R, Esmerino E, Silva M, Sant'Ana A, et al. Impact of probiotics and prebiotics on food texture. *Curr Opin Food Sci*. 2020;33:38-44. [DOI:10.1016/j.cofs.2019.12.002]
- Wieërs G, Belkhir L, Enaud R, Leclercq S, Philippart de Foy JM, Dequenne I, et al. How Probiotics Affect the Microbiota. *Front Cell Infect Microbiol*. 2019;9:454. [DOI:10.3389/fcimb.2019.00454] [PMID] [PMCID]
- Khaneghah AM, Abhari K, Eş I, Soares MB, Oliveira RB, Hosseini H, et al. Interactions between probiotics and pathogenic microorganisms in hosts and foods: A review. *Trends Food Sci Technol*. 2020;95:205-18. [DOI:10.1016/j.tifs.2019.11.022]
- Carey CM, Kostrzynska M, Ojha S, Thompson S. The effect of probiotics and organic acids on Shiga-toxin 2 gene expression in enterohemorrhagic *Escherichia coli* O157:H7. *J Microbiol Methods*. 2008;73(2):125-32. [DOI:10.1016/j.mimet.2008.01.014] [PMID]

10. Ahmadizadeh C, Mirzaei H. Evaluation of the Effect of Probiotics Isolated from Traditional Dairy Products on the Expression of Stx1 and Stx2 Genes of Verotoxigenic *Escherichia coli* (VTEC) in Laboratory Conditions. *Iran Red Crescent Med J*. 2018;20(3). [DOI:10.5812/ircmj.63544]
11. Takahashi M, Taguchi H, Yamaguchi H, Osaki T, Komatsu A, Kamiya S. The effect of probiotic treatment with *Clostridium butyricum* on enterohemorrhagic *Escherichia coli* O157:H7 infection in mice. *FEMS Immunol Med Microbiol*. 2004;41(3):219-26. [DOI:10.1016/j.femsim.2004.03.010] [PMID]
12. Asahara T, Shimizu K, Nomoto K, Hamabata T, Ozawa A, Takeda Y. Probiotic bifidobacteria protect mice from lethal infection with Shiga toxin-producing *Escherichia coli* O157:H7. *Infect Immun*. 2004;72(4):2240-7. [PMID] [PMCID] [DOI:10.1128/IAI.72.4.2240-2247.2004]
13. Wan MLY, Forsythe SJ, El-Nezami H. Probiotics interaction with foodborne pathogens: a potential alternative to antibiotics and future challenges. *Crit Rev Food Sci Nutr*. 2019;59(20):3320-33. [DOI:10.1080/10408398.2018.1490885] [PMID]
14. Cordonnier C, Thévenot J, Etienne-Mesmin L, Alric M, Livrelli V, Blanquet-Diot S. Probiotic and enterohemorrhagic *Escherichia coli*: An effective strategy against a deadly enemy?. *Crit Rev Microbiol*. 2017;43(1):116-32. [DOI:10.1080/1040841X.2016.1185602] [PMID]
15. Ku S, Yang S, Lee H, Choe D, Johnston T, Ji G, et al. Biosafety assessment of *Bifidobacterium animalis* subsp. *lactis* AD011 used for human consumption as a probiotic microorganism. *Food Control*. 2020;117:106985. [DOI:10.1016/j.foodcont.2019.106985]
16. Naylor SW, Gally DL, Low JC. Enterohaemorrhagic *E. coli* in veterinary medicine. *Int J Med Microbiol*. 2005;295(6-7):419-41. [DOI:10.1016/j.ijmm.2005.07.010] [PMID]
17. Rasko DA, Webster DR, Sahl JW, Bashir A, Boisen N, Scheutz F, et al. Origins of the *E. coli* strain causing an outbreak of hemolytic-uremic syndrome in Germany. *N Engl J Med*. 2011;365(8):709-17. [DOI:10.1056/NEJMoa1106920] [PMID] [PMCID]
18. Griffin PM, Tauxe RV. The epidemiology of infections caused by *Escherichia coli* O157:H7, other enterohemorrhagic *E. coli*, and the associated hemolytic uremic syndrome. *Epidemiol Rev*. 1991;13(1):60-98. [PMID] [DOI:10.1093/oxfordjournals.epirev.a036079]
19. Rogel Prado F, Ramón Curay R, Martínez Moreira J, Mazón Vélez J, Bayas-Morejón F. Antimicrobial activity and in vitro identification of extended-spectrum beta-lactamases (esbls) produced by *Escherichia coli* and *Klebsiella* spp. isolated from bovine mastitis cases. *J Med Pharm Chem Res*. 2024;6(6):767-79.
20. Zhao T, Doyle MP, Harmon BG, Brown CA, Mueller PO, Parks AH. Reduction of carriage of enterohemorrhagic *Escherichia coli* O157:H7 in cattle by inoculation with probiotic bacteria. *J Clin Microbiol*. 1998;36(3):641-7. [PMID] [PMCID] [DOI:10.1128/JCM.36.3.641-647.1998]
21. Abootaleb M, Mohammadi Bandari N, Karimli M, Mobayen M, Feizkhah A, Masoumi S, et al. Assessing the safety of probiotic spray as an antibiotic alternative: A clinical trial at velayat burn injuries hospital. *J Med Pharm Chem Res*. 2024;6(4):466-73.
22. Quincha Angulo A. M., Galeas Barragán, M. A., Ramón Curay, R., Mazón-Vélez, J., Bayas Morejón, F. Antimicrobial effectiveness of wild-type *Bacillus* spp vs *Bacillus subtilis* atcc 6051 against *Staphylococcus aureus* and *Klebsiella* spp isolated from bovine mastitis. *J Med Pharm Chem Res*. 2025;7(5):960-73.
23. Alhassan UMA, Abdul-Kareem IQ. Molecular identification of virulence and antimicrobial resistance genes of isolated from the patients. *J Med Pharm Chem Res*. 2025;7(4):733-44.
24. Megha G, Sumana M, Nachimuthu RP, Mahale R. Bacteriophage therapy: Unleashing the potential of bacteriophages in modern medicine. *J Med Pharm Chem Res*. 2025;7(2):282-300.
25. Van Den Bogaard AE, London N, Driessen C, Stobberingh EE. Antibiotic resistance of faecal *Escherichia coli* in poultry, poultry farmers and poultry slaughterers. *J Antimicrob Chemother*. 2001;47(6):763-71. [DOI:10.1093/jac/47.6.763] [PMID]
26. Latif Jarallah A, Fadhil Kareem A, Hassan D, Sameer Al-Rawi M, Talib Al Sa'ady A, Al-Salami I. Synthesis, Characterization, Genotoxicity Assessment and Biological Activity of Derivatives Derived from Flucytosine Drug. *Adv J Chem Section A*. 2025;8(2):245-55.
27. Shinde R, Adole V. A Anti-Microbial Evaluation, Experimental and Theoretical Insights into Molecular Structure, Electronic Properties, and Chemical Reactivity of (E)-2-((1H-indol-3-yl)methylene)-2,3-dihydro-1H-inden-1-one. *Appl Organomet Chem*. 2021;1(2): 48-58.



28. Valizadeh B, Rezaie M, Feghhi SAH, Hydarizadeh Y. R Evaluation of the Effect of UV-C on Induction of Antibiotic Resistance of *Bacillus Cereus* and *Bacillus Subtilis* Bacteria in Laboratory Conditions. *J Agric Sci Eng.* 2023;5(3):159-65.
29. Ouwehand AC, Forssten S, Hibberd AA, Lyra A, Stahl B. Probiotic approach to prevent antibiotic resistance. *Ann Med.* 2016;48(4):246-55. [DOI:10.3109/07853890.2016.1161232] [PMID]
30. Swathykrishna CS, Amrithanjali G, Shaji G, Kumar RA. Antimicrobial Activity and Synthesis of Thiazole Derivatives: A Recent Update. *J Chem Rev.* 2023;5(3):221-40.
31. Patil NA, Udgire S, Shinde DR, Patil, PD. Green Synthesis of Gold Nanoparticles using Extract of *Vitis vinifera*, *Buchananialanzan*, *Juglandaceae*, *Phoenix Dactylifera* Plants, and Evaluation of Antimicrobial Activity. *Chem Methodol.* 2023; 7(1):15-27.
32. Musa NM, Sallau MS, Oyewale AO, Ali T. Antimicrobial Activity of Lupeol and  $\beta$ -Amyrin (Triterpenoids) Isolated from the Rhizome of *Dolichos pachyrhizus* Harm. *Adv J Chem Section A.* 2024;7(1):1-14.
33. Badgujar ND, Dsouza MD, Nagargoje GR, Kadam PD, Momin KI, Bondge AS, et al. Recent Advances in Medicinal Chemistry with Benzothiazole-Based Compounds: An In-Depth Review. *J Chem Rev.* 2024;6(2):202-36.
34. Sharma P, Tomar S, Goswami P, Sangwan V, Singh R. Antibiotic resistance among commercially available probiotics. *Food Res Int.* 2014;57:176-95. [DOI:10.1016/j.foodres.2014.01.025]
35. Torabian P, Ghandehari F, Fatemi M. Biosynthesis of iron oxide nanoparticles by cytoplasmic extracts of bacteria *Lactobacillus casei*. *Asian J Green Chem.* 2018;2(3):181-8.
36. Sangchooli T, Aboulhassanzadeh S, Aghazadeh H, Paeizi M, Shokri D, Malekzadeh M. Chemical Composition, Antioxidant Properties, and Antimicrobial Activity of *Ferula assa-foetida* L. Essential Oil against Pathogenic Bacteria. *Chem Methodol.* 2024;8(5):364-385.
37. Corthésy B, Gaskins HR, Mercenier A. Cross-talk between probiotic bacteria and the host immune system. *J Nutr.* 2007;137(3):781S-90S. [DOI:10.1093/jn/137.3.781S] [PMID]
38. Das T. Ranjan Kumar A, Satpathy I, Tripathy A. Maximizing profitability and freshness: Chemical treatment techniques for live fish transportation in Odisha. *J Med Pharm Chem.* 2024;6(2):195-212.
39. Oguiche JE, Ameh AO, Bello TK, Maina NS. Samuel Prospect of Deep Eutectic Solvents in Lactic Acid Production Process: A Review. *J Chem Rev.* 2023; 5(2):96-128.
40. Kumari P, Sengar M, Sengar N. A Mini Review on Lactic Acidosis Effect, Cause, Symptoms, Complications, Metabolism, and Pathodology with Its Diagnosis, Treatment, and Preventions. *J Chem Rev.* 2024;6(1):27-38.
41. Mellies JL, Lorenzen E. Enterohemorrhagic *Escherichia coli* virulence gene regulation. In *Enterohemorrhagic Escherichia coli and Other Shiga Toxin-Producing E. coli*. 2015. pp. 175-95. Hoboken, New Jersey, United States: John Wiley & Sons, Inc. [DOI:10.1128/9781555818791.ch9] [PMID]